Parallel kinetic resolution of *tert*-butyl (*RS*)-3-alkyl–cyclopentene-1-carboxylates for the asymmetric synthesis of 3-alkyl–cispentacin derivatives

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Received 20th May 2004, Accepted 17th August 2004 First published as an Advance Article on the web 20th October 2004 OBC www.rsc.org/obc

The double mutual kinetic resolution of *tert*-butyl (*RS*)-3-benzyl–cyclopentene-1-carboxylate with a 50:50 mixture of lithium (*RS*)-*N*-benzyl-*N*- α -methylbenzylamide and lithium (*RS*)-*N*-3,4-dimethoxybenzyl-*N*- α -methylbenzylamide gives, after protonation with 2,6-di-*tert*-butylphenol, a 50:50 mixture of the readily separable *N*-benzyl-(1*SR*,2*RS*,3*RS*, α *RS*)- and *N*-3,4-dimethoxybenzyl-(1*SR*,2*RS*,3*RS*, α *RS*)- β -amino esters in >98% de in each case. This product distribution indicates that these amides react at very similar rates and with no mutual interference to furnish readily separable products, and are thus ideal for parallel kinetic resolution. The efficient parallel kinetic resolution (*E* > 65) of a range of *tert*-butyl (*RS*)-3-alkyl–cyclopentene-1-carboxylates with a pseudoenantiomeric mixture of homochiral lithium (*S*)-*N*-benzyl-*N*- α -methylbenzylamide and lithium (*R*)-*N*-3,4-dimethoxybenzyl-*N*- α -methylbenzylamide gives, after separation and *N*-deprotection, a range of carboxylate protected 3-alkyl–cispentacin derivatives in >98% de and >95% ee.

Introduction

Kinetic resolution is an underdeveloped procedure for the synthesis of enantiomerically enriched molecules.¹ While the inherent specificity of this process limits its general application within organic synthesis, the most fundamental problem with kinetic resolution is that the maximum product yield is only 50%. Furthermore, since the relative rate of a reaction depends not only upon the rate constants, but also on the relative concentrations of both reactant and substrate, as the faster reacting enantiomer is removed from a kinetic resolution reaction, the relative concentration of the slower reacting enantiomer will be augmented, and so the rate of the mismatched pairing relative to the matched pair will increase. Such concentration effects (known as mass action) influence markedly the efficiency of kinetic resolutions at conversions approaching 50%. A number of procedures have been developed to improve kinetic resolution processes, and to combat the deleterious effects of mass action as a resolution proceeds, by keeping the concentration of the reacting enantiomers identical and constant throughout the process. Dynamic kinetic resolution attempts to address these issues through the in-situ racemisation of the substrate undergoing kinetic resolution, and, provided that the rate of racemisation is faster than the rate of the disfavoured reaction, the ee of the product is independent of conversion. The result of this methodology, although rarely achieved, is that complete conversion of a racemic mixture to a single enantiomerically pure product is theoretically possible.² Alternatively, a pseudoenantiomeric mixture of two homochiral reagents may react with a racemic substrate with complementary stereoselectivities in a parallel kinetic resolution reaction to give different stereochemical, regiochemical, or differentially protected forms of the products. For parallel kinetic resolution to be viable, the pseudoenantiomeric mixture of reagents must fulfil a number of criteria:3 they must react independently without mutual interference; the reaction rate of the pseudoenantiomeric components must be similar to ensure that racemic substrate is maintained throughout the reaction; and for ease of isolation, the two diastereoisomeric products of the reaction must be readily separable or amenable to differential deprotection. For example, Vedejs et al. have demonstrated that the pseudoenantiomeric chiral DMAP equivalents 1 and 2 can effect enantioselective acyl-transfer for the parallel kinetic resolution of racemic

alcohol (*RS*)-3 (Scheme 1).⁴ While kinetic resolution of (*RS*)-3 proceeds with E = 42 using 1,⁵ the parallel kinetic resolution of (*RS*)-3 with 1 and 2 affords 5 in 46% yield and 94% ee (equivalent to E > 125) and 4 in 46% yield and 83% ee. Only recently have further efficient examples of this protocol appeared in the literature,⁶ with growing interest in this area demonstrated by a number of recent reviews.⁷



We have previously shown that the conjugate addition of homochiral lithium amides derived from α -methylbenzylamine to α,β -unsaturated esters represents an efficient and versatile approach to the asymmetric synthesis of β -amino acid derivatives.⁸ This methodology has been extended recently to the efficient kinetic resolution of a range of *tert*-butyl (*RS*)-3alkylcyclopentene-1-carboxylates,⁹ and we have demonstrated that the maximum level of enantiorecognition between the chiral α,β -unsaturated ester and chiral lithium amide in these systems may be evaluated through their mutual kinetic resolution [addition of the (*RS*)-ester to an excess of the lithium (*RS*)amide] a protocol initially suggested by Horeau.¹⁰ The effects of mass action are eliminated in this protocol, meaning that the optimal stereoselectivity factor (*E*) for the reaction is identical to the diastereoselectivity observed in the reaction, and may be calculated independent of the reaction conversion.¹¹ In order to improve the efficiency of this kinetic resolution methodology, the investigation of a pseudoenantiomeric mixture of chiral lithium amides to effect the parallel kinetic resolution of *tert*butyl (*RS*)-3-alkylcyclopentene-1-carboxylates was initiated. In order to evaluate the suitability of a range of homochiral lithium amides to act as pseudoenantiomeric resolving agents for parallel kinetic resolution, the extension of the mutual kinetic resolution strategy to a double mutual kinetic resolution protocol [addition of the (*RS*)-ester to an excess of a 50:50 mixture of lithium (*RS*)-amides] and the optimised results of this parallel kinetic resolution methodology are delineated herein (Fig. 1).¹²



Fig. 1 Kinetic and parallel kinetic resolution of 3-alkyl cyclopentene-1-carboxylates.

Results and discussion

Evaluating the efficiency of chiral lithium amides for parallel kinetic resolution: the development of a double mutual kinetic resolution protocol

Lithium (RS)-N-3,4-dimethoxybenzyl-N- α -methylbenzylamide 6, lithium (RS)-N-allyl-N- α -methylbenzylamide 7 and lithium (RS)-N-benzyl-N- α -methylbenzylamide 8, all of which show high levels of diastereoselectivity upon conjugate addition to achiral α , β -unsaturated esters, were chosen as potential reagents for parallel kinetic resolution. In the preceding manuscript, we demonstrated that high levels of enantiorecognition are observed in the mutual kinetic resolution of racemic mixtures of lithium (RS)-N-benzyl-N- α -methylbenzylamide 8 with (RS)-3-benzylcyclopentene-1-carboxylate 9 (E > 160).^{9c} Although lithium (RS)-N-3,4-dimethoxybenzylamide 6 and lithium (RS)-N-allylamide 7 may be expected to show similar levels of enantiorecognition upon conjugate addition to acceptor (RS)-9, the mutual kinetic resolution of these components was evaluated to confirm this hypothesis. In this manner, conjugate addition of lithium (RS)-N-3,4-dimethoxybenzylamide 6 to (RS)-9, employing $NH_4Cl_{(aq)}$ as an enolate protonation source, gave a mixture of three β -amino ester diastereoisomers $(1SR, 2RS, 3RS, \alpha RS)$ -10, (1*RS*,2*RS*,3*RS*,*aRS*)-11 and $(1RS, 2SR, 3SR, \alpha RS)$ -12 in a 90:9:1 ratio,¹³ with chromatographic purification giving the major diastereoisomer 10 in >98% de and in 80% isolated yield. The use of 2,6-di-tert-butylphenol as the enolate protonation source after addition of lithium (RS)-N-3,4-dimethoxybenzylamide 6 to (RS)-9 increased the C(1)-protonation selectivity, furnishing 10:11:12 in a 98:1:1 ratio, and giving (1SR,2RS,3RS,aRS)-10 in 80% isolated yield and in >98% de after chromatographic purification. The con-

figuration at C(2) within β -amino esters 10 and 11 was assigned relative to the N- α -methylbenzyl stereogenic centre by analogy to previous models rationalising the high stereoselectivity observed upon addition of homochiral lithium amides derived from α -methylbenzylamine to α , β -unsaturated esters,¹⁴ with the relative configuration at C(1) and C(3) assigned from ¹H NOE difference analysis (Scheme 2). β-Amino ester diastereoisomers 10 and 11 were shown to be epimeric at C(1), since epimerisation of the major diastereoisomer $(1SR, 2RS, 3RS, \alpha RS)$ -10 gave β -amino ester (1RS,2RS,3RS, α RS)-11 in 89% yield and >98% de over two steps via the corresponding acid, consistent with E > 99 in the conjugate addition step of the reaction. These results indicate that high levels of 'matched' anti-2,3-selectivity $(\geq 99:1)$ are observed upon addition of lithium (RS)-6 to (RS)-3-benzyl 9, with the two major diastereoisomers 10 and 11 arising from 'matched' lithium amide addition, and 11 only from loss of stereoselectivity upon enolate protonation.15



 $NR_2 = N-3,4$ -dimethoxybenzyl- $N-\alpha$ -methylbenzyl

Scheme 2 Reagents and conditions: (i) (*RS*)-6 (3 eq), THF, -78 °C then either NH₄Cl(aq) or 2,6-di-*tert*-butylphenol, THF, -78 °C to rt; (ii) 'BuOH, THF, 'BuOK, reflux; (iii) Cl₃CC=NH(O'Bu), 1:2 DCM: cyclohexane then BF₃.Et₂O.

In a similar fashion, addition of lithium (*RS*)-*N*-allylamide 7 to acceptor (*RS*)-9 and protonation of the resultant enolate with NH₄Cl gave a mixture of three β -amino ester diastereoisomers (1*SR*,2*RS*,3*RS*,*αRS*)-13, (1*RS*,2*RS*,3*RS*,*αRS*)-14 and (1*RS*,2*SR*,3*RS*,*αRS*)-15 in a 32:65:3 ratio. Exhaustive chromatographic purification yielded a mixture containing only β -amino esters 13 and 14 in a 33:67 ratio and in 85% isolated yield, which were epimerised and hydrolysed quantitatively to a single acid 16 in >98% de, consistent with 13 and 14 being epimeric at C(1). Using 2,6-di-*tert*-butylphenol as the proton source gave a 65:32:3 ratio of diastereoisomers 13:14:15, which was separated with difficulty from 2,6-di-*tert*-butylphenol by chromatography, giving a 65:32:3 mixture of 13:14:15 in 40% isolated yield. These results are consistent with high levels of 'matched' *anti*-2,3-selectivity (≥97:3) upon addition of lithium

(*RS*)-*N*-allylamide **7** to (*RS*)-3-benzyl **9**, consistent with E > 32, although inversion of the sense of selectivity in protonation of the enolate arising from conjugate addition was observed with the use of NH₄Cl and 2,6-di-*tert*-butylphenol. The relative configuration within β -amino esters **13–15** was assigned by analogy to those arising from the mutual kinetic resolution of acceptor (*RS*)-**9** with lithium amide **8** (Scheme 3).



Scheme 3 Reagents and conditions: (i) (*RS*)-7 (3 eq), THF, -78 °C then either NH₄Cl(aq) or 2,6-di-*tert*-butylphenol, THF, -78 °C to rt; (ii) 'BuOH, THF, 'BuOK, reflux.

These results demonstrate that lithium (RS)-amides 6, 7 and 8 all show high levels of enantiorecognition upon reaction with the chiral acceptor (RS)-9. However, for an efficient parallel kinetic resolution, a pseudoenantiomeric mixture of these lithium amides must react without mutual interference, at similar rates, and give separable β -amino ester products. To investigate these issues and to find the optimal mixture of lithium amides to use in a parallel kinetic resolution reaction, a double mutual kinetic resolution strategy was explored. In this reaction, acceptor (RS)-9 would be treated with a 50:50 mixture of two racemic lithium amides. It was envisaged that this competition experiment would determine the relative rates of addition of each lithium amide, while the levels of diastereoselectivity observed would probe the effects of mutual interference. The differentially protected β-amino ester products from these reactions should also be readily separated. Thus, addition of a 50:50 mixture of lithium (RS)-N-benzylamide 8 and lithium (RS)-N-3,4-dimethoxybenzylamide 6 to acceptor (RS)-9 and protonation with 2,6-di-tert-butylphenol furnished a 50:50 mixture of β -amino esters N-benzyl- $(1SR, 2RS, 3RS, \alpha RS)$ -17 in >98% de and N-3, 4-dimethoxybenzyl-(1SR,2RS,3RS, α RS)-10 in >98% de. The β -amino ester products were easily separable by chromatography, giving 17 in 28% yield and >98% de, and 10 in 26% yield and >98% de (Scheme 4).

Addition of a 50:50 mixture of lithium (*RS*)-*N*-allylamide 7 and lithium (*RS*)-*N*-benzylamide 8 to acceptor (*RS*)-9 gave a 45:22:33 mixture of *N*-allyl β -amino esters 13:14 to *N*-benzyl β -amino ester 17. This product distribution indicates that although conjugate addition occurs *anti*- to the C(3)-stereodirecting group with each lithium amide, low levels of selectivity are observed upon protonation of the intermediate *N*-allyl- β -amino ester enolate, as observed in the mutual kinetic resolution protocol. Thus, although lithium amides 7 and 8 show no mutual interference in their conjugate addition, the



Scheme 4 Reagents and conditions: (i) (RS)-6 (1.5 eq), (RS)-8 (1.5 eq), THF, -78 °C to rt; (ii). 2,6-di-*tert*-butylphenol, THF, -78 °C to rt.

rate of addition of lithium N-allylamide 7 is twice the rate of addition of lithium N-benzylamide 8. Chromatography facilitated separation of the N-allyl and N-benzyl β-amino ester products, giving the inseparable N-allyl β-amino esters 13:14 in 33% de and 15% yield and N-benzyl β-amino ester 17 in >98% de and 18% yield. Consistent with the observed rate difference, the (RS)-acceptor was treated with a 67:33 mixture of lithium N-benzylamide 8 (2 eq) and lithium N-allylamide 7 (1 eq), generating a 33.5:16.5:50 mixture of N-allyl β -amino esters 13:14 to N-benzyl β-amino ester 17. As expected, addition of a 50:50 mixture of lithium (RS)-N-allylamide 7 and lithium (RS)-N-3,4-dimethoxybenzylamide 6 to (RS)-9 gave a 45:22:33 mixture of N-allyl β-amino esters 13:14 to N-3,4dimethoxybenzyl β -amino ester 10, with chromatographic purification giving the inseparable N-allyl β -amino esters 13:14 in 33% de and 13% yield, and N-benzyl β -amino ester 10 in >98% de and 12% yield (Scheme 5).

These results are consistent with the mixtures of lithium amides *N*-benzyl-**8**, *N*-3,4-dimethoxybenzyl-**6** and *N*-allyl-**7** showing no mutual interference in their reactions. The rates of the addition of lithium amides *N*-benzyl-**8** and *N*-3,4-dimethoxybenzyl-**6** are essentially identical, and half that of *N*-allyl-**7**. For simplicity, it was decided that the parallel kinetic resolution of 3-alkyl–cyclopentene-1-carboxylates would be undertaken using 50:50 mixtures of homochiral lithium (*S*)-*N*-benzylamide **8** and lithium (*R*)-*N*-3,4-dimethoxybenzylamide **6**.

Parallel kinetic resolution of (*RS*)-3-alkyl–cyclopentene-1carboxylates with a pseudoenantiomeric mixture of lithium (*R*)-*N*-3,4-dimethoxybenzyl-*N*-α-methylbenzylamide and lithium (*S*)-*N*-benzyl-*N*-α-methylbenzylamide

Addition of a 50:50 mixture of lithium (*R*)-*N*-3,4-dimethoxybenzylamide **6** (>98% ee)¹⁶ and lithium (*S*)-*N*-benzylamide **8** (>98% ee) to acceptor (*RS*)-**9** gave a 50:50 mixture of β -amino esters (1*S*,2*R*,3*R*,*aR*)-**10** and (1*R*,2*S*,3*S*,*aS*)-**17** in >98% de in each case, consistent with *E* > 70. β -Amino esters **10** and **17** were readily separable by chromatography, giving (1*S*,2*R*,3*R*,*aR*)-**10** in 36% yield and >98% de and (1*R*,2*S*,3*S*,*aS*)-**17** in 40% yield and >98% de (Scheme 6).

To demonstrate the efficiency and generality of this methodology, the parallel kinetic resolution of the known (RS)-3-methyl and (RS)-3-ethyl acceptors **18** and **19** was examined.¹⁷ Addition of a 50:50 mixture of lithium (R)-*N*-3,4-dimethoxybenzylamide **6** and lithium (S)-*N*-benzylamide **8** to (RS)-3-methyl **18** gave a 50:50 mixture of β -amino esters



Scheme 5 Reagents and conditions: (i) (*RS*)-7 (1.5 eq), (*RS*)-8 (1.5 eq), THF, -78 °C; (ii) (*RS*)-6 (1.5 eq), (*RS*)-7 (1.5 eq), THF, -78 °C; (iii) 2,6-di-*tert*-butylphenol, THF, -78 °C to rt.



Scheme 6 Reagents and conditions: (i) (*R*)-6 (1.5 eq), (*S*)-8 (1.5 eq), THF, -78 °C; (ii) 2,6-di-*tert*-butylphenol, THF, -78 °C to rt.

N-3,4-dimethoxybenzyl **20**: *N*-benzyl **21**, each in 98 \pm 1% de, consistent with *E* > 65 in each case. Chromatographic purification gave the *N*-3,4-dimethoxybenzyl- β -amino ester **20** in 40% yield (>98% de), and the *N*-benzyl- β -amino ester **21** in 34% yield (>98% de). Similarly, addition of a 50:50 mixture of lithium amides (*R*)-**6**:(*S*)-**8** to (*RS*)-3-ethyl **19** gave a 50:50 mixture of β -amino esters *N*-3,4-dimethoxybenzyl **22**:*N*-benzyl **23**, each in 98 \pm 1% de (*E* > 65), with purification giving **22** in 30% yield (>98% de) and **23** in 34% yield (Scheme 7).

Deprotection: Synthesis of the enantiomers of 3-alkylcispentacin derivatives

With a range of pseudoenantiomeric β -amino esters prepared by parallel kinetic resolution, *N*-deprotection to the



 20, R=Me, 40%, >98% d.e.
 21, R=Me, 34%, >98% d.e.

 22, R=Et, 30%, >98% d.e.
 23, R=Et, 34%, >98% d.e.

Scheme 7 Reagents and conditions: (i) (*R*)-6 (1.5 eq), (*S*)-8 (1.5 eq), THF, -78 °C; (ii) 2,6-di-*tert*-butylphenol, THF, -78 °C to rt.

corresponding enantiomeric primary β-amino esters was investigated. Hydrogenolysis of the 3-benzyl, 3-methyl and 3-ethyl-N-benzyl-N-α-methylbenzyl protected β-amino esters 17, 21, and 23 gave the corresponding primary β -amino esters (1R,2S,3S)-24, (1R,2S,3R)-25 and (1R,2S,3R)-26, respectively, in good yields and in >98% de. The ee of each β -amino ester was shown to be >95% ee by derivatisation with both racemic and 99% ee Mosher's acid chloride and comparison of the 19F and ¹H NMR spectra of the resulting amides (Scheme 8). In the enantiomeric series, N-deprotection of the 3-benzyl, 3-methyl and 3-ethyl-N-3,4-dimethoxybenzyl-N-a-methylbenzyl β-amino esters 10, 22, and 23 to the corresponding primary β -amino esters was effected using a two-step deprotection protocol, as attempted global N-deprotection by hydrogenolysis proved problematic. Thus, oxidative removal of the N-3,4-dimethoxybenzyl protecting group within 10, 22 and 23 with DDQ gave the corresponding *N*- α -methylbenzyl protected β -amino esters 27-29 in good yield and >98% de, with subsequent hydrogenolysis of N- α -methylbenzyl protected β -amino esters 27–29 giving the β -amino esters (1S,2R,3R)-24, (1S,2R,3S)-25 and (1*S*,2*R*,3*S*)-**26** in >98% de and >95% ee.

In conclusion, initial screening has determined that homochiral lithium (*S*)-*N*-benzyl-*N*- α -methylbenzylamide **8** and (*R*)-*N*-3,4-dimethoxybenzyl-*N*- α -methylbenzylamide **6** constitute an effective pseudoenantiomeric mixture for the parallel kinetic resolution of 3-alkyl cyclopentene-1-carboxylates, giving a 50:50 mixture of β -amino ester products in >98% de in each case. The *N*-benzyl and *N*-3,4-dimethoxybenzyl β -amino ester products are readily separated by chromatographic purification, affording differentially protected 3-substituted cispentacin derivatives in high yield and >98% de in all cases examined, which are easily *N*-deprotected to the corresponding primary β amino esters. The extension of this methodology to the parallel kinetic resolution of 5-alkyl–cyclopentene-1-carboxylates and a range of substituted cyclohexene-1-carboxylates is currently under investigation within our laboratory.

Experimental

General experimental

All reactions were carried out under nitrogen or argon using standard vacuum line techniques and glassware that was flame dried and cooled under nitrogen. THF was distilled from sodium/benzophenone ketyl; *n*-butyllithium was used as a solution in hexanes and was titrated against diphenylacetic



Scheme 8 Reagents and conditions: (i) $Pd(OH)_2$ on C, MeOH, H_2 (5 atm), rt; (ii) DDQ (2.1 eq), DCM : H_2O (3:1), rt.

acid prior to use. All other reagents were used as supplied without further purification. Flash column chromatography was performed on silica gel (Kieselgel 60). TLC was performed on Merck aluminium sheets coated with 0.2 mm silica gel 60 F₂₅₄. Plates were visualised either by UV light (254 nm), iodine, ammonium molybdate (7% solution in ethanol), potassium permanganate (1% in 2% aqueous acetic acid, containing 7% potassium carbonate), or Dragendorff's reagent.¹⁸ Infrared spectra were recorded as thin films or KBr discs using a Perkin-Elmer PARAGON 1000 FT-IR spectrometer, with selected peaks reported in cm⁻¹. ¹H and ¹³C NMR spectra were recorded on Varian Gemini 200 (1H 200 MHz, 13C 50 MHz), Bruker DPX-200 (1H 200 MHz, 13C 50 MHz), Bruker DPX-400 and AVANCE AV-400 (1H 400 MHz, 13C 100 MHz), or Bruker AMX-500 (1H 500 MHz, 13C 125 MHz) spectrometers. Chemical shifts ($\delta_{\rm H}$) are reported in parts per million (ppm) and are referenced to the residual solvent peak, with coupling constants (J) measured in hertz. Low resolution mass spectra (m/z) were recorded on either a VG Masslab 20–250 instrument (CI, NH₃) or Platform instrument (ESI). Major peaks are listed with intensities quoted as percentages of the base peak. Accurate mass measurements were recorded on a VG Autospec and a Waters 2790-Micromass LCT electrospray ionisation mass spectrometer operating at a resolution of 5000 full-width halfheight. Positive ion spectra were calibrated relative to PEG with tetraoctylammonium bromide as the internal lock mass. Negative ion spectra were calibrated relative to poly(DL-alanine) with leucine enkephalin as the internal lock mass. Optical rotations were recorded on a Perkin-Elmer 241 polarimeter, using a path length of 10 cm, in spectroscopic grade solvents (Aldrich), with concentrations (c) given in g per 100 cm^3 , solvent and temperature as recorded. Melting points were recorded on a Gallenkamp Hot Stage apparatus and are uncorrected. Elemental analyses were performed by the microanalysis service of the Inorganic Chemistry Laboratory, University of Oxford.

General procedure 1: Lithium amide additions to α , β -unsaturated esters. *n*-BuLi was added dropwise to a stirred solution of amine in THF at -78 °C. After 30 min, the ester in THF at -78 °C was added *via* cannula and stirred for 2 h before the addition of either: (method A) aqueous NH₄Cl, the resultant mixture extracted with DCM, and the organic layer concentrated *in vacuo*. The residue was redissolved in DCM, washed successively with 10% aqueous citric acid, aqueous NaHCO₃, and brine, dried, filtered, and concentrated *in vacuo*; or (method B) addition of 2,6-di-*tert*-butylphenol (DTBP) in THF *via* cannula, stirred for 30 min at -78 °C, and then warmed to 0 °C and stirred for a further 30 min before the solvent was removed *in vacuo*.

General procedure 2: Epimerization of β -amino acid derivatives. A catalytic amount of 'BuOK was added to the β -amino ester in a 1:1 solution of 'BuOH and THF. The solution was refluxed for 18 h before addition of excess aqueous NH₄Cl, separation and extraction with DCM. The combined organic phases were dried, filtered, and concentrated *in vacuo*.

General procedure 3: Hydrogenolysis. Pd(OH)₂/C was added to a solution of secondary or tertiary amine in degassed MeOH at room temperature and placed under a hydrogen atmosphere (5 atm). After stirring for 24 h, the reaction mixture was filtered through basic alumina, washed with MeOH and concentrated *in vacuo.*

General procedure 4: Deprotection of *N*-3,4-dimethoxybenzylamine derivatives. The adduct was dissolved in 5:1 DCM: water, and DDQ was added. The reaction was stirred at rt for 2 days, before addition of aqueous NaHCO₃, extraction with DCM, and washing with brine. Combined organic phases were dried, filtered, and concentrated *in vacuo*.

Preparation of *tert*-butyl (1SR,2RS,3RS,αRS)-3-benzyl-2-(N-3,4-dimethoxybenzyl-N-α-methylbenzylamino)cyclopentane-1-carboxylate 10

Following General Procedure 1, n-BuLi (0.91 mL, 2.21 mmol), (RS)-6 (617 mg, 2.28 mmol), and (RS)-9 (200 mg, 0.76 mmol) in THF (10 mL), quenching with DTBP (460 mg, 2.21 mmol) gave, after purification by chromatography (3% Et₂O in pentane), 10 (320 mg, 80%) as a colourless oil; $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.05–1.15 (1H, m, C(5)H_A), 1.25 (3H, d, J 7.1, C(α)CH₃), 1.49 $(9H, s, C(CH_3)_3), 1.49-1.51 (1H, m, C(5)H_B), 1.68-1.82 (2H, m)$ C(4)*H*₂), 2.08 (1H, dd, *J* 3.0, 1.1, C*H*_AH_BPh), 2.47–2.52 (1H, m, C(3)H), 2.56–2.61 (1H, m, C(1)H), 3.07–3.11 (1H, m, C(2)H), $3.42 (1H, dd, J 3.4, 1.5, CH_A H_B Ph), 3.89 (3H, s, C_6 H_3 (OCH_3)_A),$ 3.91 (3H, s, C₆H₃(OCH₃)_B), 3.99 (1H, d, J 15.6, CH_AH_BAr), 4.22 (1H, d, J 15.6, CH_AH_BAr), 4.25 (1H, q, J 7.0, C(α)H), 6.88–7.74 (13H, m, *Ph*, *Ar*); $\delta_{\rm C}(100 \text{ MHz}, \text{CDCl}_3)$ 20.9 (C(α)*C*H₃), 27.2 (C(4)), 28.1 (C(5)), 28.1 (OC(CH₃)₃), 41.0 (CH₂Ph), 42.2 (C(3)), 46.6 (C(1)), 50.5 (CH₂Ar), 55.7, 55.9 (C₆H₃ (OCH₃)₂), 60.3 $(C(\alpha)), 69.1 (C(2)), 80.0 (OC(CH_3)_3), 110.7, 119.3, 125.6, 126.9,$ 127.6, 128.1, 128.3, 128.6, 135.9, 145.3, 147.5 (Ph, Ar), 175.8 (C=O); $v_{\text{max}}/\text{cm}^{-1}$ (film) 1716 (C=O); m/z (ESI⁺) 530 (MH⁺, 100%); HRMS found 530.3272, C₃₄H₄₄NO₄ requires 530.3270.

Preparation of *tert*-butyl (1*RS*,2*RS*,3*RS*,α*RS*)-3-benzyl-2-(*N*-3,4-dimethoxybenzyl-*N*-α-methylbenzylamino)cyclopentane-1-carboxylate 11

Following *General Procedure 2*, **10** (50 mg, 0.11 mmol), 'BuOH (10 mL), THF (5 mL) and 'BuOK gave a carboxylic acid (44.0 mg, 0.12 mmol, 100%), which was then dissolved in a 2:1 cyclohexane: DCM (10 mL) with Cl₃CC=NH(O'Bu) (48 mg, 0.24 mmol).¹⁹ The mixture was stirred for 10 h, then BF₃-Et₂O (2 μ L, 0.08 mmol) was added, followed by saturated aqueous NaHCO₃ after 1 h. The aqueous layer was extracted into DCM and the combined organic phases were dried and concentrated *in vacuo*. Purification by chromatography (10% Et₂O in petrol)

gave 11 (45 mg, 89%) as a colourless oil; $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.26–1.33 (1H, m, C(5) H_A), 1.27 (3H, d, J 4.2, C(α)C H_3), 1.47 (9H, s, $C(CH_3)_3$), 1.48–1.53 (1H, m, $C(5)H_B$), 1.65–1.77 (2H, m, C(4)H₂), 1.96–2.23 (1H, m, C(3)H), 2.14–2.25 (1H, m, CH_AH_BPh), 2.74–2.78 (1H, m, C(1)H), 3.22–3.27 (1H, m, CH_AH_BPh), 3.45 (1H, dd, J 4.2, 1.6, C(2)H), 3.64 (1H, d, J 15.8, CH_AH_BAr), 3.72 (1H, d, J 15.8, CH_AH_BAr), 3.89 (3H, s, C₆H₃(OCH₃)_A), 3.87–3.93 (1H, q, J 5.5 C(α)H), 3.90 (3H, s, $C_6H_3(OCH_3)_B$, 6.86–7.40 (13H, m, *Ph*, *Ar*); $\delta_C(100 \text{ MHz}, \text{CDCl}_3)$ 21.3 (C(a)CH₃), 28.1 (OC(CH₃)₃), 29.7 (C(4)), 30.2 (C(5)), 40.1 (CH₂Ph), 45.4 (C(1)), 47.0 (C(3)), 50.2 (CH₂Ar), 55.7, 56.0 $(C_6H_3(OCH_3)_2), 61.6(C(\alpha)), 70.0(C(2)), 77.0(OC(CH_3)_3), 110.8,$ 111.0, 119.2, 125.6, 126.7, 127.6, 127.8, 128.2, 128.6, 128.8, 135.6, 141.9, 145.2, 148.9 (*Ph*, *Ar*), 176.2 (*C*=O); *v*_{max}/cm⁻¹ (film) 1720 (C=O, s); m/z (ESI+) 530 (MH+, 100%); HRMS found 530.3279, C₃₄H₄₄NO₄ requires 530.3270.

Preparation of *tert*-butyl (1*SR*,2*RS*,3*RS*, α *RS*)-3-benzyl-2-(*N*-allyl-*N*- α -methylbenzylamino)cyclopentane-1-carboxylate 13 and *tert*-butyl (1*RS*,2*RS*,3*RS*, α *RS*)-3-benzyl-2-(*N*-allyl-*N*- α -methylbenzylamino)cyclopentane-1-carboxylate 14

Following General Procedure 1, n-BuLi (0.91 mL, 2.27 mmol), (RS)-7 (0.38 mg, 2.28 mmol), and (RS)-9 (200 mg, 0.76 mmol) in THF (6 mL), quenching with saturated aqueous NH₄Cl, gave after purification by chromatography (1% Et₂O in pentane), a 33:67 mixture of 13:14 (270 mg, 85%) as a yellow oil. $(1SR, 2RS, 3RS, \alpha RS)$ -13: $\delta_{H}(400 \text{ MHz}, \text{CDCl}_{3})$ 1.37 (3H, d, J 7.1, C(a)CH₃), 1.42-1.51 (2H, m, C(4)H₂), 1.49 (9H, s, OC(CH₃)₃), 1.70-1.73 (2H, m, C(5)H₂), 2.01-2.06 (1H, m, C(3)H), 2.15-2.21 (1H, m, CH_AH_BPh), 2.63-2.70 (1H, m, C(1)H), 3.18 (1H, dd, J 9.9, 3.5, CH_AH_BPh), 3.28-3.30 (2H, m, NCH₂), 3.31–3.34 (1H, m, C(2)H), 3.94–3.99 (1H, q, J 7.1, $C(\alpha)H$ 5.09–5.13 (1H, m, CH=CH_AH_B), 5.13–5.36 (1H, m, $CH=CH_AH_B$, 5.91–6.00 (1H, m, $CH=CH_2$), 7.12–7.50 (10H, m, *Ph*); δ_C(100 MHz, CDCl₃) 21.6 (C(*a*)*C*H₃), 28.1 (OC(*C*H₃)_β), 29.5 (C(4)), 30.8 (C(5)), 40.0 (CH₂Ph), 45.9 (C(1)), 46.9 (C(3)), 49.3 (NCH₂), 60.8 (C(α)), 70.1 (C(2)), 77.0 (OC(CH₃)₃), 114.8 (CH=*C*H₂), 125.6, 126.6, 127.6, 127.7, 128.2, 128.4, 128.7, 128.8, 142.3 (*Ph*_{o,m,p}), 140.0 (*C*H=CH₂), 145.2, 145.8 (*Ph*_{inso}), 176.4 (C=O); v_{max}/cm^{-1} (film) 1732 (C=O); m/z (ESI⁺) 420 (MH+, 100%); HRMS found 420.2898, C₂₈H₃₇NO₂ requires 420.2903. (1RS,2RS,3RS, α RS)-14: $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.38 (3H, d, J 6.8, C(α)CH₃), 1.49 (9H, s, C(CH₃)₃), 1.54–1.79 (4H, m, C(4)H₂, C(5)H₂), 1.98 (1H, dd, J 11.1, 2.3, CH_AH_BPh), 2.35-2.55 (2H, m, C(1)H, C(3)H), 2.92 (1H, dd, J 8.3, 2.0, C(2)H), 3.19 (1H, d, J 9.9, 3.5, CH_AH_BPh), 3.50 (1H, ddt, J 10.9, 5.3, 1.8, CH_AH_BCH=CH₂), 3.63 (1H, ddt, J 10.3, 5.8, 1.5, $CH_AH_BCH=CH_2$, 4.25 (1H, q, J 6.8, C(α)H), 5.04–5.14 (1H, m, $CH = CH_AH_B$, 5.24–5.35 (1H, m, $CH = CH_AH_B$), 5.87–6.04 (1H, m, CH=CH₂), 7.05-7.50 (10H, m, Ph).

Preparation of (1*RS*,2*RS*,3*RS*,α*RS*)-3-benzyl-2-(*N*-allyl-*N*-α-methylbenzylamino)cyclopentane-1-carboxylic acid 16

Following General Procedure 2, a mixture of 13:14 (50 mg, 0.11 mmol), 'BuOH (20 mL), THF (20 ml) and 'BuOK gave 16 (43.7 mg, 100%) as an off-white solid; $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.39 (3H, d, J 7.1, C(α)CH₃), 1.41-1.53 (2H, m, C(5)H₂), 1.49 (9H, s, C(CH₃)₃), 1.70–1.73 (2H, m, C(4)H₂), 2.01–2.06 (1H, m, C(3)H), 2.15-2.21 (1H, m, CH_AH_BPh), 2.65-2.70 (1H, m, C(1)H), 3.18 (1H, dd, J 6.0, 2.2, CH_AH_BPh), 3.35–3.43 (2H, m, NCH₂), 3.31–3.34 (1H, m, C(2)H), 4.10 (1H, q, J 4.2, C(α)H), 5.11 (1H, q, J 10.6, 1.8, CH=C H_AH_B), 5.29 (1H, q, J 17.2, 1.8, CH=CH_AH_B), 5.86–5.96 (1H, m, CH=CH₂), 7.12–7.56 (10H, m, Ph); $\delta_{C}(100 \text{ MHz}, \text{CDCl}_{3})$ 25.1 (C(α)CH₃), 32.1 (C(4)), 33.0 (C(5)), 43.6 (CH₂Ph), 48.2 (C(1)), 50.1 (C(3)), 53.1 (NCH₂), 65.6 (*C*(α)), 74.3 (*C*(2)), 118.5 (CH=*C*H₂), 118.5, 129.3, 130.6, 131.4, 131.9, 132.2, 142.9, 145.7 (Ph), 146.1 (CH=CH₂), 180.0 (C=O); v_{max}/cm⁻¹ (KBr) 3435 (OH), 1690 (C=O); m/z (ESI⁻) 361 (M-H-, 52%); HRMS found 364.2287, C₂₈H₃₂NO₂ requires 364.2277; mp 100-104 °C.

Double mutual kinetic resolution for the preparation of *tert*butyl (1*SR*,2*RS*,3*RS*,*αRS*)-3-benzyl-2-(*N*-benzyl-*N*-*α*-methylbenzylamino)cyclopentane-1-carboxylate 17 and *tert*-butyl (1*SR*,2*RS*,3*RS*,*αRS*)-3-benzyl-2-(*N*-3,4-dimethoxybenzyl-*N*-*α*methylbenzylamino)cyclopentane-1-carboxylate 10

Following General Procedure 1, (RS)-9 (100 mg, 0.39 mmol), (RS)-8 (124 mg, 0.59 mmol), (RS)-6 (159 mg, 0.59 mmol) and *n*-BuLi (0.47 mL, 1.18 mmol) in THF (5 mL), quenching with DTBP (244 mg, 1.18 mmol) gave, after purification by chromatography (0.5% Et₂O in petrol), **17** (51 mg, 28%) and **10** (53 mg, 26%) with identical properties to those reported below and above, respectively.

Double mutual kinetic resolution for the preparation of *tert*-butyl (1*SR*,2*RS*,3*RS*,*aRS*)-3-benzyl-2-(*N*-benzyl-*N*-*a*-methylbenzylamino)cyclopentane-1-carboxylate 17, *tert*-butyl (1*SR*,2*RS*,3*RS*,*aRS*)-3-benzyl-2-(*N*-allyl-*N*-*a*-methyl-benzylamino)cyclopentane-1-carboxylate 13 and *tert*-butyl (1*RS*,2*RS*,3*RS*,*aRS*)-3-benzyl-2-(*N*-allyl-*N*-*a*-methylbenzylamino)cyclopentane-1-carboxylate 13 and *tert*-butyl (1*RS*,2*RS*,3*RS*,*aRS*)-3-benzyl-2-(*N*-allyl-*N*-*a*-methylbenzylamino)cyclopentane-1-carboxylate 13 and *tert*-butyl (1*RS*,2*RS*,3*RS*,*aRS*)-3-benzyl-2-(*N*-allyl-*N*-*a*-methylbenzyl-

Following General Procedure 1, (RS)-9 (100 mg, 0.39 mmol), (RS)-8 (124 mg, 0.59 mmol), (RS)-7 (94 mg, 0.59 mmol) and *n*-BuLi (0.47 mL, 1.18 mmol) in THF (5 mL), quenching with DTBP (244 mg, 1.18 mmol) gave, after purification by chromatography (0.5% Et₂O in petrol), 17 (32 mg, 18%) and 13:14 (24 mg, 15%) with identical properties to those reported below and above, respectively.

Double mutual kinetic resolution for the preparation of *tert*butyl (1*SR*,2*RS*,3*RS*, α *RS*)-3-benzyl-2-(*N*-3,4-dimethoxybenzyl-*N*- α -methylbenzylamino)cyclopentane-1-carboxylate 10 and *tert*-butyl (1*SR*,2*RS*,3*RS*, α *RS*)-3-benzyl-2-(*N*-allyl-*N*- α -met hylbenzylamino)cyclopentane-1-carboxylate 13 and *tert*-butyl (1*RS*,2*RS*,3*RS*, α *RS*)-3-benzyl-2-(*N*-allyl-*N*- α -methylbenzylamino)cyclopentane-1-carboxylate 14

Following General Procedure 1, (RS)-9 (100 mg, 0.39 mmol), (RS)-6 (159 mg, 0.59 mmol), (RS)-7 (94 mg, 0.59 mmol) and *n*-BuLi (0.47 mL, 1.18 mmol) in THF (5 mL), quenching with DTBP (244 mg, 1.18 mmol) gave, after purification by chromatography (0.5% Et₂O in petrol), **10** (19 mg, 12%) and **13**:14 (24 mg, 13%) with identical properties to those reported above.

Parallel kinetic resolution for the preparation of *tert*-butyl $(1R,2S,3S,\alpha S)$ -3-benzyl-2-(N-benzyl-N- α -methylbenzylamino)-cyclopentane-1-carboxylate 17 and *tert*-butyl $(1S,2R,3R,\alpha R)$ -3-benzyl-2-(N-3,4-dimethoxybenzyl-N- α -methyl benzylamino)cyclopentane-1-carboxylate 10

Following General Procedure 1, (RS)-9 (300 mg, 1.16 mmol), (S)-8 (379 mg, 1.80 mmol), (R)-6 (488 mg, 1.80 mmol), and n-BuLi (1.39 mL, 3.47 mmol) in THF (5 mL), quenching with DTBP (716 mg, 3.49 mmol) gave, after purification by chromatography (0.5% ether in petrol), **17** (218 mg, 40%) as a colourless solid; δ_H(400 MHz, CDCl₃) 1.03-1.12 (1H, m, C(4)H_A), 1.25 (3H, d, J 6.8, C(α)CH₃), 1.41–1.49 (1H, m, C(5)H_A), 1.45 (9H, s, C(CH₃)₃), 1.65–1.79 (2H, m, C(4)H_B, C(5)H_B), 2.05 (1H, dd J 13.7, 11.4, CH_AH_BPh), 2.42-2.45 (1H, m, C(3)H), 2.56 (1H, ddd, J 7.8, 4.6, 3.6, C(1)H), 3.06 (1H, dd, J 10.3, 7.8, C(2)H), 3.31 (1H, dd, J 13.4, 3.0, CH_AH_BPh), 4.08 (1H, d, J 16.2, CH_AH_BPh), 4.33 (1H, d, J 16.2, CH_AH_BPh), 4.27 (1H, q, J 6.8, C(α)H), 7.14–7.55 (15H, m, Ph); $[a]_{D}^{24}$ –46.3 (c 0.73 in CHCl₃) (lit.,^{9c} [a]²⁴_D-45.1 (c 0.49 in CHCl₃)); mp 96–98 °C (lit., 9° 86–88 °C). Further elution yielded 10 (214 mg, 36%) with identical spectroscopic properties to those reported above and $[a]_{D}^{24}$ +75.2 (*c* 0.70 in CHCl₃).

Parallel kinetic resolution for the preparation of *tert*-butyl (1*R*,2*S*,3*R*,α*S*)-3-methyl-2-(*N*-benzyl-*N*-α-methylbenzylamino)-cyclopentane-1-carboxylate 21 and *tert*-butyl

(1*S*,2*R*,3*S*,α*R*)-3-methyl-2-(*N*-3,4-dimethoxybenzyl-*N*-α-methyl benzylamino)cyclopentane-1-carboxylate 20

Following General Procedure 1, (RS)-18 (300 mg, 1.64 mmol), (S)-8 (519 mg, 2.46 mmol), (R)-6 (664 mg, 2.46 mmol), and n-BuLi (2.36 mL, 5.92 mmol) in THF (10 mL), quenching with DTBP (1.21 g, 5.91 mmol) gave, after purification by chromatography (0.5% ether in petrol) 21 (219 mg, 34%) as a colourless oil; $\delta_{\rm H}(400 \text{ MHz}, \text{CDCl}_3) 0.99-1.04 (1\text{H}, \text{m}, \text{C}(4)H_{\rm A})$, 1.02 (3H, d, J 6.5, C(3)CH₃), 1.19 (3H, d, J 6.9, C(α)CH₃), 1.51 (9H, s, OC(CH₃)₃), 1.50–1.54 (1H, m, C(5)H_A), 1.68–1.76 (1H, m, C(5) $H_{\rm B}$), 1.95–1.99 (1H, m, C(4) $H_{\rm B}$), 2.24–2.32 (1H, m, C(3)H), 2.53 (1H, ddd, J 7.7, 7.7, 4.0, C(1)H), 2.82 (1H, dd, J 10.5, 7.7, C(2)H), 3.96 (1H, d, J 15.9, CH_AH_BPh), 4.16 (1H, d, J 15.9, CH_AH_BPh), 4.15 (1H, q, J 6.9, C(α)H), 7.21–7.52 (10H, m, Ph). Further elution yielded 20 (297 mg, 40%) as a colourless oil; (Found: C, 73.7; H, 8.2; N, 3.1. C₂₈H₄₀NO₄ requires C, 74.1; H, 8.7; N, 3.1%); $\delta_{\rm H}(400 \text{ MHz, CDCl}_3) 0.93$ (3H, d, J 6.5, C(3)CH₃), 0.96–1.02 (1H, m, C(4)H_A), 1.22 (3H, d, J 6.7, C(a)CH₃), 1.50 (9H, s, OC(CH₃)₃), 1.54–1.58 (1H, m, C(5)H_A), 1.68–1.80 (1H, m, C(5)H_B), 1.93–2.06 (1H, m, C(4)H_B), 2.21-2.37 (1H, m, C(3)H), 2.39-2.58 (1H, m, C(1)H), 2.84 (1H, dd, J 7.5, 2.9, C(2)H), 3.87 (1H, d, J 15.1, CH_AH_BAr), 3.89 (3H, s, C₆H₃(OCH₃)_{*A*}), 3.94 (3H, s, C₆H₃(OCH₃)_{*B*}), 4.09 (1H, d, *J* 15.1, CH_A*H*_BAr), 4.16 (1H, q, *J* 6.8, C(α)*H*), 6.58–7.65 (8H, m, *Ph*, Ar); δ_C(100 MHz, CDCl₃) 19.7 (C(3)CH₃), 20.1 (C(α)CH₃), 27.0 (C(4)), 28.1 $(OC(CH_3)_3)$, 30.7 (C(5)), 34.6 (C(3)), 47.1 (C(1)), 50.5 (CH₂Ar), 55.8, 55.9 (C₆H₃(OCH₃)₂), 59.6 (C(a)), 70.3 (C(2)), 79.9 (C(CH₃)₃), 110.8, 111.0, 119.3, 126.7, 127.7, 127.9, 128.0, 128.2, 135.8, 145.3, 147.4, 148.8 (Ar, Ph), 175.9 (C=O); v_{max} /cm⁻¹ (film) 1717 (C=O); $[a]_{D}^{24}$ +75.3 (c 0.70 in CHCl₃); m/z (ESI+) 454 (MH+, 100%).

Parallel kinetic resolution for the preparation of *tert*-butyl (1*R*,2*S*,3*R*, α *S*)-3-ethyl-2-(*N*-benzyl-*N*- α -methylbenzylamino)-cyclopentane-1-carboxylate 23 and *tert*-butyl (1*S*,2*R*,3*S*, α *R*)-3-ethyl-2-(*N*-3,4-dimethoxybenzyl-*N*- α -methylbenzylamino)cyclopentane-1-carboxylate 22

Following General Procedure 1, (RS)-19 (300 mg, 1.53 mmol), (S)-8 (484 mg, 2.30 mmol), (R)-6 (626 mg, 2.30 mmol), and n-BuLi (3.6 mL, 4.60 mmol) in THF (10 mL), quenching with DTBP (923 mg, 4.60 mmol) gave, after purification by chromatography (0.5% ether in petrol), 23 (219 mg, 34%) as a colourless oil; $\delta_{\rm H}(400 \text{ MHz}, \text{CDCl}_3) 0.82 (3\text{H}, \text{t}, J 7.3, \text{C}(3)\text{CH}_2\text{CH}_3)$, 0.94–0.99 (1H, m, C(3)CH_AH_BCH₃), 1.00–1.07 (1H, m, C(4)H_A), 1.19 (3H, d, J 6.8, $C(\alpha)CH_3$), 1.49–1.51 (1H, m, $C(5)H_A$), 1.52 $(9H, s, OC(CH_3)_3), 1.70-1.75 (1H, m, C(5)H_B), 1.88-1.90 (1H, m)$ m, C(3)CH_AH_BCH₃), 1.95–2.03 (1H, m, C(4)H_B), 2.04–2.07 (1H, m, C(3)H), 2.54 (1H, ddd, J7.6, 7.6, 4.0, C(1)H), 2.92 (1H, dd, J 10.0, 7.6, C(2)H), 3.94 (1H, d, J 15.9, CH_AH_BPh), 4.17 (1H, d, J15.9, CH_AH_BPh), 4.16 (1H, q, J 6.8, C(α)H), 7.21–7.48 (10H, m, *Ph*); $[a]_D^{24} - 126.3$ (*c* 0.70 in CHCl₃) (lit., ^{9c} $[a]_D^{24} - 125.3$ (c 1.11 in CHCl₃)). Further elution yielded 22 (214 mg, 30%) as a colourless oil; $\delta_{\rm H}(400 \text{ MHz}, \text{CDCl}_3) 0.85 (3\text{H}, \text{t}, J 7.3,$ C(3)CH₂CH₃), 0.92–1.00 (1H, m, C(3)CH_AH_BCH₃), 1.02–1.11 (1H, m, C(4)H_A), 1.21 (3H, d, J 7.2, C(a)CH₃), 1.51 (9H, s, C(CH₃)₃), 1.52-1.55 (1H, m, C(5)H_A), 1.70-1.74 (1H, m, C(5)H_B), 1.81–1.92 (1H, m, C(3)CH_AH_BCH₃), 1.98–2.15 (2H, m, C(3)H, C(4)H_B), 2.49–2.54 (1H, m, C(1)H), 2.91–2.95 (1H, dd, J 7.8, 2.3, C(2)H), 3.80 (1H, d, J 15.5, CHAHBAr), 3.89 $(3H, s, C_6H_3(OCH_3)_A), 3.93 (3H, s, C_6H_3(OCH_3)_B), 4.08 (1H, d, d)$ J 15.5, CH_A H_B Ar), 4.12 (1H, q, J 3.2, C(α)H), 6.79–7.44 (8H, m, *Ph*, *Ar*); δ_C(100 MHz, CDCl₃) 12.6 (CH₂CH₃), 22.0 (C(α)CH₃), 27.5 (CH₂CH₃), 28.1 (OC(CH₃)₃) 29.3 (C(4)), 30.4 (C(5)), 45.8 $(C(1)), 46.6(C(3)), 50.3(CH_2DMP), 55.8, 55.9(C_6H_3(OCH_3)_2),$ 61.6 (*C*(α)), 68.4 (*C*(2)), 77.6 (O*C*(CH₃)₃), 110.8, 119.1, 126.7, 127.6, 128.2, 136.1, 145.4, 147.4, 148.8 (*Ph*, *Ar*), 176.4 (*C*=O); v_{max} /cm⁻¹ (film) 1717 (C=O); $[a]_{D}^{24}$ +30.1 (c 0.70 in CHCl₃); m/z (ESI⁺) 468 (MH⁺, 100%); HRMS found 468.3114, C₂₉H₄₂NO₄ requires 468.3115.

Preparation of *tert*-butyl (1*R*,2*S*,3*S*)-3-benzyl-2aminocyclopentane-1-carboxylate 24

Following *General Procedure 3*, **17** (200 mg, 0.43 mmol) and Pd(OH)₂/C (30 mg) under H₂ (5 atm) in MeOH (4 mL) gave (1*R*,2*S*,3*S*)-**24** (88 mg, 75%) as a colourless oil; $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.19–1.24 (1H, m, C(4)*H*_A), 1.48 (9H, s, OC(C*H*₃)₃), 1.60 (2H, br s, N*H*₂), 1.82–1.95 (3H, m, C(4)*H*_B, C(5)*H*₂), 2.06 (1H, m, C(3)*H*), 2.48 (1H, dd, *J* 13.5, 8.8, C*H*_AH_BPh), 2.84–2.91 (2H, m, CH_AH_BPh, C(1)*H*), 3.13 (1H, dd, *J* 7.4, 7.3, C(2)*H*), 7.12–7.45 (5H, m, *Ph*); $[a]_{\rm D}^{\rm 24}$ –34.0 (*c* 1.20 in CHCl₃), (lit.,^{9c} $[a]_{\rm D}^{\rm 23}$ –32.5 (*c* 1.00 in CHCl₃)).

Preparation of *tert*-butyl (1*R*,2*S*,3*R*)-3-methyl-2aminocyclopentane-1-carboxylate 25

Following *General Procedure 3*, **21** (53 mg, 0.13 mmol) and Pd(OH)₂/C (10 mg) under H₂ (5 atm) in MeOH (10 mL) gave, after purification by chromatography (20% ether in petrol), (1*R*,2*S*,3*R*)-**25** (12 mg, 45%) as a colourless oil; $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.01 (3H, d, *J* 6.4, C(3)CH₃), 1.12–1.15 (1H, m, C(4)H_A), 1.40 (9H, s, OC(CH₃)₃), 1.80 (2H, br s, NH₂), 1.83–1.97 (4H, m, C(3)H, C(4)H_B, C(5)H₂), 2.82–2.88 (1H, m, C(1)H), 2.94 (1H, dd, *J* 7.4, 7.4, C(2)H); $[a]_{\rm D}^{24}$ –55.1 (*c* 0.70 in CHCl₃) (lit.,^{9b} $[a]_{\rm D}^{24}$ –51.8 (*c* 1.02 in CHCl₃)).

Preparation of (1*R*,2*S*,3*R*)-*tert*-butyl 3-ethyl-2aminocyclopentane-1-carboxylate 26

Following *General Procedure 3*, **23** (200 mg, 0.5 mmol) and Pd(OH)₂/C (20 mg) under H₂ in MeOH (10 mL) gave (1*R*,2*S*,3*R*)-**26** (62 mg, 60%) as a yellow oil; $\delta_{\rm H}$ (400 MHz, CDCl₃) 0.90 (3H, t, *J* 7.2, CH₂CH₃), 1.09–1.23 (2H, m, (C(4)H_A, C(3)CH_AH_BCH₃), 1.45 (9H, s, OC(CH₃)₃), 1.59–169 (2H, m, C(3)H, C(4)H_B), 1.80–1.97 (5H, m, C(5)H₂, C(3)CH_AH_BCH₃, NH₂), 2.76 (1H, ddd, *J* 7.5, 7.5 and 7.9, C(1)H), 3.05–3.11 (1H, m, C(2)H); [a]_D²⁴ –46.5 (*c* 0.70 in CHCl₃) (lit.,^{9c} [a]_D²³ –44.7 (*c* 1.00 in CHCl₃)).

Preparation of *tert*-butyl (1*S*,2*R*,3*R*,α*R*)-3-benzyl-2-(*N*-α-methylbenzylamino)cyclopentane-1-carboxylate 27

Following General Procedure 4, 10 (70 mg, 0.13 mmol) and DDQ (41 mg, 0.26 mmol) in 5:1 DCM:water (20 mL) gave, after purification by chromatography (30% ether in petrol), 27 (33 mg, 70%) as a yellow oil; (Found: C, 79.4; H, 8.7; N, 3.7. C₂₅H₃₄NO₂ requires C, 79.1; H, 8.9; N, 3.7%); $\delta_{\rm H}(400~{\rm MHz},$ $CDCl_3$) 0.98–1.13 (1H, m, C(4) H_A), 1.15–1.20 (1H, m, C(5) H_A), 1.30 (3H, d, J 20.3, C(α)CH₃), 1.53 (9H, s, OC(CH₃)₃), 1.61 (1H, br s, NH), 1.63–1.83 (2H, m, C(4)H_B, C(5)H_B), 2.07–2.19 (2H, m, CH_AH_BPh, C(3)H), 2.60 (1H, dd, J 6.9, 2.0, C(2)H), 2.89–3.00 (1H, m, C(1)H), 3.10 (1H, q, J 8.6, CH_AH_BPh), 3.88 (1H, q, J 6.4, C(α)H), 7.05–7.45 (10H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃) 26.7 (C(5)), 28.2 (OC(CH_3)₃), 28.7 (C(4)), 25.4 (C(α)C H_3), 39.6 (CH₂Ph), 46.4 (C(3)), 46.7 (C(1)), 56.4 (C(a)), 64.3 (C(2)), 80.1 $(C(CH_3)_3)$, 125.5, 126.9, 127.0, 128.1, 128.3, 128.9 $(Ph_{o,m,p})$, 141.6 (Ph_{ipso}) , 176.2 (C=O); v_{max}/cm^{-1} (film) 1717 (C=O); $[a]_{D}^{24}$ +115.1 (c 0.70 in CHCl₃); m/z (ESI⁺) 380 (MH⁺, 100%); HRMS found 380.2590, C₂₅H₃₄NO₂ requires 380.2590.

Preparation of *tert*-butyl (1*S*,2*R*,3*S*,α*R*)-3-methyl-2-(*N*-α-methylbenzylamino)cyclopentane-1-carboxylate 28

Following *General Procedure* 4, **20** (200 mg, 0.44 mmol) and DDQ (160 mg, 0.88 mmol) in 5:1 DCM:water mixture (20 mL) gave, after purification by chromatography (20% Et₂O in petrol), **28** (100 mg, 75%) as a colourless oil; $\delta_{\rm H}(400 \text{ MHz, CDCl}_3)$ 0.95 (3H, d, *J* 6.1, C(3)*CH*₃), 0.97–1.08 (1H, m, C(5)*H*_A), 1.29 (3H, d, *J* 6.8, C(a)*CH*₃), 1.53 (9H, s, OC(*CH*₃),) 1.66 (1H, br s, N*H*), 1.67–1.93 (4H, m, C(3)*H*, C(4)*H*₂, C(5)*H*_B), 2.40 (1H, dd, *J* 7.9, 1.7, C(1)*H*), 2.84–2.95 (1H, m, C(2)*H*), 3.91 (1H, q, *J* 4.2, C(a)*H*), 7.15–7.44 (5H, m, *Ph*); $\delta_{\rm C}(100 \text{ MHz, CDCl}_3)$ 18.1 (C(3)*CH*₃), 25.5 (C(a)*CH*₃), 26.7 (*C*(4)), 28.2 (OC(*CH*₃)₃), 31.3

(*C*(5)), 39.2 (*C*(3)), 48.8 (*C*(1)), 56.3 (*C*(α)), 66.1 (*C*(2)), 80.0 (O*C*(CH₃)₃), 126.8, 126.8, 128.3 (*Ph*_{o,m,p}), 146.0 (*Ph*_{ipso}), 175.1 (*C*=O). v_{max}/cm⁻¹ (film) 3333 (NH), 1718 (C=O); [a]₂²⁴ +172.2 (*c* 0.70 in CHCl₃); *m*/*z* (ESI⁺) 304 (MH⁺, 100%); HRMS found 304.2271, C₁₉H₃₀NO₂ requires 304.2277.

Preparation of *tert*-butyl (1*S*,2*R*,3*S*,α*R*)-3-ethyl-2-(*N*-α-methylbenzylamino)cyclopentane-1-carboxylate 29

Following General Procedure 4, 22 (200 mg, 0.42 mmol) and DDQ (190 mg, 0.84 mmol) in 5:1 DCM:water (10 mL) gave, after purification by chromatography (20% ether in petrol), 29 (94 mg, 70%) as a colourless oil; $\delta_{\rm H}$ (400 MHz, CDCl₃) 0.85 (3H, t, J 7.1, CH₂CH₃), 0.88–1.10 (2H, m, C(3)H_AH_BCH₃, C(4)H_A), 1.28 (3H, d, J 6.3, C(α)CH₃), 1.53 (9H, s, OC(CH₃)₃), 1.53 (1H, br s, NH), 1.63–1.77 (3H, m, C(4)H_B, C(5)H_A, C(3)CH_AH_BCH₃), 1.76-1.98 (2H, m, C(3)H, C(5)H_B), 2.50 (1H, dd, J 7.7, 1.8, C(1)*H*), 2.86–2.92 (1H, m, C(2)*H*), 3.89 (1H, q, J 5.5, C(α)*H*), 7.19-7.70 (5H, m, Ph); δ_C(100 MHz, CDCl₃) 12.4 (CH₂CH₃), 25.5 (C(α)CH₃), 26.4 (C(3)CH₂CH₃), 26.7 (C(5)), 28.5 (C(CH₃)₃), 29.4 (C(4)), 46.3 (C(3), 47.1 (C(1)), 56.3 (C(a)), 64.4 (C(2)), 80.0 (C(CH₃)), 126.7, 126.8, 128.0, 128.3 (Ph_{o,m,p}), 146.0 (Ph_{ipso}) , 175.0 (C=O); v_{max} /cm⁻¹ (film) 1718 (C=O); $[a]_D^{24}$ +124.0 (c 0.70 in CHCl₃); m/z (ESI⁺) 318 (MH⁺, 100%); HRMS found 318.2432, C₂₀H₃₂NO₂ requires 318.2433.

Preparation of *tert*-butyl (1*S*,2*R*,3*R*)-3-benzyl-2aminocyclopentane-1-carboxylate 24

Following *General Procedure 3*, **27** (200 mg, 0.53 mmol) and Pd(OH)₂/C (30 mg) under H₂ (5 atm) in MeOH (4 mL) gave (1*S*,2*R*,3*R*)-**24** (65 mg, 45%) with ¹H NMR data identical to those reported above; $[a]_{2}^{2b}$ +36.0 (*c* 1.10 in CHCl₃).

Preparation of *tert*-butyl (1*S*,2*R*,3*S*)-3-methyl-2aminocyclopentane-1-carboxylate 25

Following *General Procedure 3*, **28** (50.0 mg, 0.17 mmol) and Pd(OH)₂/C (10 mg) under H₂ (5 atm) in MeOH (10 mL) gave (1*S*,2*R*,3*S*)-**25** (11.8 mg, 35%) as a colourless oil with ¹H NMR data identical to those reported above; $[a]_D^{24}$ +54.1 (*c* 0.50 in CHCl₃).

Preparation of *tert*-butyl (1*S*,2*R*,3*S*)-3-ethyl-2aminocyclopentane-1-carboxylate 26

Following *General Procedure 3*, **29** (150 mg, 0.47 mmol) and Pd(OH)₂/C (70 mg) under H₂ (5 atm) in MeOH (5 mL) gave (1*S*,2*R*,3*S*)-**26** (70 mg, 70%) as a yellow oil with ¹H NMR data identical to those reported above; $[a]_{D}^{24}$ +40.3 (*c* 0.7 in CHCl₃).

Acknowledgements

The authors wish to thank Pfizer for an industrial CASE award (J.M.W.), the Rhodes Trust (M.J.S.) and New College, Oxford for a Junior Research Fellowship (A.D.S.)

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